DENITRIFIER ENZYME ACTIVITY

D.J. Herman

Materials

225 mL jelly jars, with lids fitted with Hungate septa Glucose KNO $_3$ Acetylene 10 mL disposable plastic syringes N_2 Shaker

Preparation

- 1. 1 mM Glucose 1 mM KNO₃ solution: Dissolve 0.180 g glucose and 0.101 g KNO₃ to 1,000 mL in a volumetric flask.
- 2. Purify the acetylene by passing the gas through a solution of CuCl₂ in concentrated HCl and then water.

Procedure

- 1. Place 50 g soil and 30 mL glucose-KNO₃ solution in jelly jar, and seal with a fresh lid. Shake vigorously.
- 2. Include 2 control jars, with 30 mL glucose-KNO₃ solution but no soil.
- 3. Alternately evacuate and flush with N_2 four times. Equilibrate with atmospheric pressure after the last N_2 flush using a glass syringe.
- 4. Remove 20 mL of the headspace and replace with 20 mL purified acetylene.
- 5. Shake the flask vigorously, then incubate on a rotary shaker (about 100 rpm).
- 6. Draw at least 4 syringe samples during an incubation period not to exceed 2 h. For each injection, add 3 mL of room air, pump the syringe 3 times with headspace, then draw the 3 mL sample.

References

- Tiedje, J.M. 1982. Denitrification. pp 1011-1026. IN: Page, A.L., R.H. Miller, and D.R. Keeney (eds). Methods of soil analysis. Part 2. Second edition. American Society of Agronomy, Madison, WI, USA.
- 2. Tiedje, J.M. 1994. Denitrifiers. pp 245-267. *IN*: Weaver, R.W., J.S. Angle, and P.S. Bottomley (eds). Methods of soil analysis. Part 2. American Society of Agronomy, Madison, WI, USA.