Removing DNA from DNA/RNA mixtures:

Reagents:

10 X digestion buffer (20 mM $MgCl_2$, 10 mM $ZnCl_2$, 500 mM Tris, pH 7.2)
DNase I
Protein precipitation solution (10 M ammonium acetate)
Ice-cold Isopropanol
Ice-cold Ethanol (70 %)
DEPC treated water or 1 x TE buffer

1. Add 1/10 volume of 10 X digestion buffer and 1 U of DNase I, mix by flicking tubes and spin briefly (do not vortex this will reduce the activity of DNase).

2. Incubate at 37^{o}C for 30 min.

3. To remove enzyme add 1 volume of protein precipitation solution, vortex for 10s on low speed and place in ice-bath for 5 min.

4. Centrifuge at top speed (16,100 x g) at 4^{o}C for 10 min and transfer supernatant to a new tube.

5. Add 1 volume of ice-cold isopropanol and invert tube about 30 times to mix.

6. Centrifuge at top speed (16,100 x g) at 4^{o}C for 10 min and remove supernatant.

7. Rinse pellet twice with 70 % ice-cold ethanol spinning between rinses.

8. Dry pellet slightly in vacuum dessicator (about 2 min).

9. Resuspend pellet either in sterile H_2O or in 1 X TE buffer.

10. Store at -20^{o}C short term or -80^{o}C long term.